

patient's permanent record. For preferred results, it is recommended that the end of the extent of both responses be recorded. This can be accomplished by using the longest erythema diameter, then selecting the mid-point of that line and a line at right angles to that line to determine the orthogonal diameter. The sum of these two diameters is the sum of wheal diameters (ΣE); the sum of wheal diameters is determined as follows:

Dilution	Extract	Diluent	BAU/mL
0	Concentrate		100,000
1	0.5 mL concentrate	4.5 mL	10,000
2	0.5 mL dilution 1	4.5 mL	1,000
3	0.5 mL dilution 2	4.5 mL	100
4	0.5 mL dilution 3	4.5 mL	10
5	0.5 mL dilution 4	4.5 mL	1
6	0.5 mL dilution 5	4.5 mL	0.1

*Due to differences such as source material, preservative, potency dilutions, storage conditions, and length of storage, there is no common potency correlation ratio between extracts standardized in Bioequivalent Allergy Units (BAU) and:

- 1) standardized extracts previously labeled in Allergy Units (AU);
- 2) non-standardized extracts labeled weight-to-volume (w/v);
- 3) non-standardized extracts labeled in Protein Nitrogen Units (PNU); or
- 4) alum-precipitated extracts.

Stock mixtures of grass pollen extracts are compounded from individual grass pollen extracts. The total potency per milliliter (mL) of these mixtures is described in DOSAGE FORMS, COMPOSITION AND PACKING.

OVERDOSAGE

Signs and symptoms of overdose are typically local and systemic reactions. For a description and management of overdose reactions, refer to "Adverse Reactions" section above.

ACTION AND CLINICAL PHARMACOLOGY

Mechanism of Action

Diagnostically (for skin testing), the allergen combines with IgE antibodies fixed to mast cells in the skin. This complexing causes an increase in cellular permeability and degranulation of the mast cells releasing chemical mediators. These mediators (such as histamine) are responsible for a local inflammatory response of wheal and erythema typical of a positive skin test reaction and also, the symptoms commonly associated with allergic disease. The more mediator released, the larger the reaction (wheal and erythema).

Treatment consists of the subcutaneous injection of gradually increasing doses of the allergens to which the patient is allergic. It has been demonstrated that this method of treatment induces an increased tolerance to the allergens responsible for the symptoms on subsequent exposure. Although the exact relationship between allergen, skin-sensitizing antibody (IgE) and the blocking antibody (IgG) has not been precisely established, clinically confirmed immunological studies have adduced evidence of the efficacy of hyposensitization therapy.

Numerous controlled studies have demonstrated the clinical efficacy of immunotherapy with cat, dust mites and some pollens, including grass pollen extracts.⁴ Nevertheless, responses are variable, and in a few studies patients reported no appreciable benefit.

Puncture test data with 10,000 BAU/mL Grass Pollen Extract CBER reference preparations, in 15 grass allergic patients yielded the following sizes of wheal and erythema (Σ = sum of longest diameter and orthogonal cross diameter) >

Table 1: Puncture bifurcated needle data with 10,000 BAU/mL CBER Reference Grass Pollen Extracts.

Reference Pollen	FDA Lot #	N	P _Σ Erythema (mm)		P _Σ Wheal (mm)	
			Mean	Range	Mean	Range
Bermuda	E4-Ber	15	90.3	43-123	15.7	7-31
June	E3-Jkb	15	77.3	47-107	15.9	6-28
Meadow Fescue	E4-MF	15	81.1	57-115	11.9	7-22
Orchard	E4-Or	15	84.3	57-111	14.1	9-19
Perennial Rye	E10-Rye	15	92.3	73-135	17.5	6-36
Redtop	E4-Rt	15	77.1	42-98	14.1	8-19
Sweet Vernal	E4-SV	15	81.2	28-123	15.7	8-30
Timothy	E6-Ti	15	88.3	51-109	16.9	8-40

The intradermal dose (BAU₅₀) of the CBER (FDA) Grass Pollen Extract Reference Preparation required to produce a 50 mm Sum of Erythema was calculated based on titration in sensitive individuals.

Table 2: Intradermal Dose of CBER Reference Grass Pollen Extracts for 50 mm Sum of Erythema Diameter (BAU₅₀)⁵.

Reference Pollen	FDA Lot #	BAU ₅₀ /mL	
		Mean	Range
Bermuda	E4-Ber	0.02	0.4-0.0003
June	E3-Jkb	0.02	0.1-0.004
Meadow Fescue	E4-MF	0.02	0.9-0.002
Orchard	E4-Or	0.02	1.9-0.002
Perennial Rye	E10-Rye	0.02	0.7-0.002
Redtop	E4-Rt	0.02	0.8-0.004
Sweet Vernal	E4-SV	0.02	1.0-0.002
Timothy	E6-Ti	0.02	0.6-0.002

An analysis of relative potency of the 1:10 w/v unstandardized grass pollen extracts utilizing the ELISA Inhibition method shows the relative potency in BAU/mL in the following table. **CAUTION:** By the very nature of unstandardized extracts, individual lots of the unstandardized extracts may vary more than 1 log from the average value expressed in these tables.

TABLE 3: Estimation of Potency Described in BAU/mL by ELISA-Inhibition of ALK Abello, Inc. 1:10 w/v Non-standardized Grass Pollen Extracts Manufactured and Distributed by ALK-Abello, Inc. (Formerly Center Laboratories, Inc.)

1:10 w/v Extract	# Lots Assayed	Ave PNU/mL	Range PNU/mL	Estimated BAU/mL	Range * BAU/mL	BAU/PNU RATIO

Physicians must exercise care in switching patients from non-standardized to standardized extracts. As with non-standardized extracts, dosage with BAU extracts must be derived based on the patient's sensitivity to the specific pollen. Switching from an extract that was not standardized in BAU cannot be made by a calculated, numerical ratio, but TABLE 3 can be used as a guide. Dose selection can be confirmed by side-by-side testing of non-standardized and standardized extracts at estimated equal doses. See **WARNINGS** section. Patients being switched from non-standardized extracts from another manufacturer to extracts standardized in BAU can be re-evaluated by diagnostic skin testing to judge the dose to start immunotherapy or to build up to new maintenance dosages.

Duration of Effect

The usual duration of treatment has not been established. A period of two or three years of injection therapy constitutes an average minimum course of treatment. Patients should have sufficient treatments before each pollen season.

STORAGE AND STABILITY

To maintain stability of allergenic extracts, proper storage conditions are essential. Bulk concentrates and diluted extracts are to be stored at 2° to 8° C even during use. Bulk or diluted extracts are not to be frozen. Do not use after the expiration date shown on the vial label.

SPECIAL HANDLING INSTRUCTIONS

Clinicians should be aware that diluted extracts are inherently less stable than concentrates. Dilutions of glycerinated extracts which result in glycerin below 50% may also be less stable. Potency of a particular dilution can be checked by skin test in comparison to a fresh dilution of the extract on an individual known to be allergic to the specific antigen.

DOSAGE FORMS, COMPOSITION AND PACKAGING

For percutaneous testing, 5 mL vial, 100,000 BAU/mL in glycerin 50% (v/v).

For immunotherapy, 10 mL and 50 mL vials 100,000 BAU/mL in glycerin 50% (v/v).

Composition of Standardized Grasses in each product:

Individual standardized grass pollen includes 100,000 BAU/mL of either one of the following grasses: June (*Poa pratensis*), Meadow Fescue (*Festuca elatior*), Orchard (*Dactylis glomerata*), Perennial Rye (*Lolium perenne*), Redtop (*Agrostis alba*), Sweet Vernal (*Anthoxanthum odoratum*), and Timothy (*Phleum pratense*).

Mixture of 4 standardized grass pollen:

June (<i>Poa pratensis</i>)	25,000 BAU/mL
Orchard (<i>Dactylis glomerata</i>)	25,000 BAU/mL
Redtop (<i>Agrostis alba</i>)	25,000 BAU/mL
Timothy (<i>Phleum pratense</i>).	25,000 BAU/mL

Mixture of 5 standardized grass pollen:

June (<i>Poa pratensis</i>)	20,000 BAU/mL
Orchard (<i>Dactylis glomerata</i>)	20,000 BAU/mL
Redtop (<i>Agrostis alba</i>)	20,000 BAU/mL
Sweet Vernal (<i>Anthoxanthum odoratum</i>)	20,000 BAU/mL
Timothy (<i>Phleum pratense</i>).	20,000 BAU/mL

PHARMACEUTICAL INFORMATION

Drug Substance

Proper name:	Standardized Timothy Grass Standardized Orchard Grass Standardized June Grass Standardized Redtop Standardized Sweet Vernal Standardized Perennial Rye Grass Standardized Meadow Fescue Grass
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Product Characteristics

Standardized allergenic extract of grass pollens from Timothy (*Phleum pratense*), Orchard (*Dactylis glomerata*), June (*Poa pratensis*), Redtop (*Agrostis alba*), Sweet Vernal (*Anthoxanthum odoratum*), Meadow Fescue (*Festuca elatior*), Perennial Rye (*Lolium perenne*), mixture of 4 Standard grass pollen (Timothy, Orchard, June and Redtop) and mixture of 5 Standard grass pollen (Timothy, Orchard, June, Redtop and Sweet Vernal) in the accompanying vial are sterile, and contain glycerine 50% v/v and phenol 0.4% (preservative). Inert ingredients may include sodium chloride for isotonicity and sodium bicarbonate buffer.

DETAILED PHARMACOLOGY

The allergic reaction is dependent upon the presence of antigen-specific immunoglobulin E (IgE) antibodies that are bound to specific receptors on mast cells and basophils. The presence of IgE antibodies on mast cells and basophils sensitizes these cells and--upon interaction with the appropriate allergen--histamine and other mediators are released. IgE antibody has been shown to correlate with atopic diseases such as allergic rhinitis and allergic asthma. In the skin these mediators are responsible for the characteristic wheal and flare (erythema) reactions upon allergenic extract skin testing in persons with the specific allergies.

Specific immunotherapy with pollen extracts as employed for many years is helpful in reducing symptoms associated with exposure to the offending allergens. A summary of effectiveness by the Panel on Review of Allergenic Extracts, an advisory committee to the U. S. Food and Drug Administration, has been published. Several mechanisms have been proposed to explain the effectiveness of immunotherapy; an increase in antigen-specific IgG antibodies is frequently associated with clinical effectiveness, although correlation is not consistent in all studies; there is a decrease in specific IgE; and IgE production is suppressed during periods of seasonal or high exposure to the antigen. Other changes following immunotherapy have been noted including development of auto-anti-idiotypic antibodies, a decrease in blood basophil sensitivity to allergen, a decrease in lymphokine production and lymphocyte proliferation by cells exposed to allergen, and development of allergen-specific suppressor cells. The complete mechanisms of immunotherapy are not known and remain the subject of investigation.

REFERENCES

	Reference
1	Turkeltaub, P. C. <i>et al.</i> Office of Biologics Research and Review skin test method for evaluation of subject sensitivity to standardized allergenic extracts and for assignment of allergy units to reference preparations using the ID50/EAL method. FDA CBER Methods of the Allergenic Products Testing Laboratory 1993
2	Miller, CA; Boyle, KT and Braun, M. ELISA Competition Assay - Quantitative Determination of Relative Potency of Allergenic Extracts. FDA CBER Methods of the Allergenic Products Testing Laboratory. 1993
3	Norman, P. S. The clinical significance of IgE. <i>Hosp. Prac.</i> 1975; 10:41-49.
4	VanMette, T. E. and Adkinson, N. F. Immunotherapy for aeroallergen disease. In: Middleton <i>et al.</i> Allergy Principles and Practice 3rd Ed. St. Louis: CV Mosby, 1988:1327.
5	Data on file at FDA
6	Umetzu, D. T. <i>et al.</i> Serum sickness triggered by anaphylaxis: a complication of immunotherapy. <i>J. Allergy Clin. Immunol.</i> 1985; 76:713.
7	Phanphanhak, P. and Kohler, P. F. Onset of polyarteritis nodosa during allergic hyposensitization treatment. <i>Am. J. Med.</i> 1980; 68:479.
8	Köhler, P. F. Immune complexes and allergic disease. In: Middleton <i>et al.</i> Allergy Principles and Practice 3rd Ed. St. Louis: CV Mosby, 1988:167.
9	Bousquet, J. In vivo methods for the study of allergy: skin test, techniques, and interpretation. In: Middleton <i>et al.</i> Allergy Principles and Practice 3rd Ed. St. Louis: CV Mosby, 1988:167.
10	Committee on the Safety of Medicines. CSM update: desensitizing vaccines. <i>Brit. Med. J.</i> 1986; 293:948.
11	Lockey, R. F. <i>et al.</i> Fatalities from immunotherapy (IT) and skin testing (ST). <i>J. Allergy Clin. Immunol.</i> 1987; 79:660.
12	Reid, M. J. <i>et al.</i> Survey of fatalities from skin testing and immunotherapy 1985-1989. <i>J. Allergy Clin. Immunol.</i> 1993; 92:6.
13	DuBuske L. M. <i>et al.</i> Special problems regarding Allergen immunotherapy in Immunology and Allergy Clinics of North America, Greenburger, PA. Ed. February 1992; 145-149.
14	Freedman, SO. Asthma and Allergic Rhinitis II Clinical Aspects. In: Freedman and Gold Clinical Immunology, 2nd Ed. Hagerstown, MD: Harper & Row, 1976; 131.
15	Saetan N, Rhyre MB, Mellits ED, et al. Immunotherapy of pollenosis in children: investigation of the immunologic basis of clinical improvement. <i>N Eng J Med</i> 1969;280:623.
16	Johnstone DE: Value of hyposensitization therapy for perennial bronchial asthma in children. <i>Pediatrics</i> 1961;27:39.
17	VanAsperin PP, Kemp AS, Mellis CM: Skin test reactivity and clinical allergen sensitivity in infancy. <i>J Allergy Clin Immunol</i> 1994;73:381-6.

to begin at 1/10 of the dose that produces sum of erythema of 50 mm (2+ positive skin test reaction). For example, if a patient exhibits a 2+ on to 1 BAU/mL, the first dose should be no higher than 0.05 mL of 0.1 mL may be increased by 0.05 mL each time until 0.5 mL is reached, at which old more concentrated dilution can be used, beginning with 0.05 mL, if no is observed.

of an allergic extract has been established, the initial dose from the new reduced to 25% of the previously well tolerated dose (see also **WARNINGS** and **CONTRAINDICATIONS**).

doses in the early stages of immunotherapy is no more than once to twice gradually be increased to once every two weeks. Generally, maintenance given as infrequently as once every two weeks to once a month.

subcutaneously preferably in the arm. It is advantageous to give alternate arms and routinely in the same area. In some patients, a local allergic reaction may develop thus preventing a possible severe local reaction.

needle, but before injecting the dose, pull plunger of the syringe slightly. If the syringe, discard the syringe and contents and repeat injection at another site.

extracts must be diluted for initial therapy and intradermal skin testing. For subsequent therapy, refer to DOSAGE AND ADMINISTRATION section.

slowly become less potent with age. During the course of treatment, if a patient continues to have a reaction to an extract bearing a later expiration date, the extract bearing the later expiration date should be lowered to a safe level. When switching one standardized extract with another, at least one dilution is suggested.

Precautions when making dilutions. The first dose of the new extract should be at least 75% of the amount of the dosage from the previous extract.

or diluted forms of this product are not complete. The undiluted product will under recommended storage conditions at least until the expiration date on each vial. It is recommended that minimal amounts of the concentrate be used. Diluted product is used up within a relatively short period of time; i.e., within four weeks.

Grass Pollen Allergenic Extracts are supplied as sterile solutions for subcutaneous administration.

For intradermal testing, use of Sterile Diluent for Allergenic Extracts or Sterile Diluent for